

Oestrogen receptors in breast cancer: basic mechanisms and clinical implications

Cecilia Williams and Chin-Yo Lin

Center for Nuclear Receptors and Cell Signaling, Department of Biology and Biochemistry, University of Houston, Houston, Texas 77204, USA

Correspondence to: Chin-Yo Lin. Email: clin23@central.uh.edu

Abstract

Since the discovery of the connection between ovarian hormones and breast cancer, endocrine therapy has been an integral adjuvant treatment for patients with hormone-dependent breast cancers. Oestrogen receptor (ER) plays a central role in mediating the effects of endogenous hormones and therapeutic agents. ER serves as a prognostic marker for responsiveness to endocrine therapy and is targeted either directly by selective oestrogen receptor modulators (SERMs) and pure antagonists or indirectly by aromatase inhibitors (AIs) that block oestrogen production. A significant number of ER-positive patients, however, fail to respond to therapy or develop resistance over time. This review focuses on the current understanding of ER functions and recent advances in genomic technologies and research that have provided a global perspective on hormone and ER activity and led to a number of significant discoveries, including the roles of co-regulatory factors and non-coding RNAs. Mechanistic insights into normal ER functions and therapeutic actions of SERMs and AIs will enable the development of better predictive markers and more effective target mechanisms and ultimately facilitate improvements in disease outcomes and patient survival.

Keywords: *breast cancer, hormonal carcinogenesis, endocrine therapy, oestrogen receptor*

Published: 05/11/2013

Received: 15/08/2013

ecancer 2013, 7:370 DOI: 10.3332/ecancer.2013.370

Copyright: © the authors; licensee ecancermedicalsecience. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

*A lady with growth neoplastic
thought castration was just a bit drastic.
She preferred that her ill could be cured with a pill.
Today it's no longer fantastic.*

This quatrain, composed by Elwood Jensen and V. Craig Jordan, amusingly and succinctly summarises one of the great triumphs in breast cancer research and treatment [1]. In 1896, George Beatson reported the beneficial effects of oophorectomy, the female equivalent of castration, in two of his patients with inoperable breast cancer [2]. From his earlier studies of ovulation and lactation, Beatson astutely made the connection between ovarian functions and influences, subsequently shown to be the ovarian hormone oestrogen, with phenotypic changes in mammary tissues and possible link to cancer. He took the first steps in testing this hypothesis, and his seminal discovery provided the first evidence for hormonal carcinogenesis and the potential efficacy of targeting ovarian and hormonal functions. With contributions by Jensen, Jordan, and many others, endocrine therapy, using pills that block oestrogen production or activity, is now routinely applied in the treatment of breast cancer. Other examples of targeted therapy in breast cancer include the use of monoclonal antibodies (trastuzumab) and small molecule receptor tyrosine kinase inhibitors (lapatinib) in targeting the HER2/neu growth factor receptor-positive tumours [3]. This review focuses on the role and mechanisms of action of oestrogen receptors (ERs) in mediating the effects of oestrogen and endocrine therapeutic agents and discusses current challenges and opportunities in targeting ER and oestrogen signalling in the prevention and treatment of breast cancer.

Discovery and characterisation of ERs

Jensen and Jacobson were the first to observe the retention of radiolabelled oestrogen in hormone-responsive target tissues [4]. Subsequently, work by Jensen, Gorski, and their respective groups demonstrated the existence of intracellular oestrogen-binding receptor proteins [5–8]. The *ER* gene was cloned by the Chambon group, and mutagenesis studies showed that the receptor consists of a DNA-binding domain containing zinc finger motifs and a ligand-binding domain, key structural elements of ligand-dependent transcription factors [9, 10]. Functional studies also identified the N-terminal activating function (AF-1) domain, which is involved in protein–protein interactions important for the transcriptional activity of ER [11]. The identification of other related receptors places ER in the nuclear receptor superfamily of transcriptional regulators [12]. Molecular characterisation of ER revealed that, upon ligand activation, ER regulates target gene expression by binding cis-regulatory elements termed oestrogen response elements (EREs; consensus 5'-GGTCAnnnTGACC-3'). This interaction is facilitated by the pioneering factor FOXA1 [13]. ER can also bind DNA indirectly by tethering to other transcription factors, including AP-1, Sp1, NFκB, and RUNX1. DNA-bound ER nucleates co-regulator complexes that modify chromatin and render the DNA accessible to the transcriptional machinery [14, 15]. ER co-regulators include those that enhance transcriptional activity by altering nucleosome spatial orientation (SWI/SNF) or by modifying histones through acetylation (SRC1, CBP/p300, p/CAF, and p/CIP/AIB1) and methylation (CARM1, PRMT1) [16–25]. Some co-regulators such as NCoR, SMRT, NRIP1, LCoR, and REA function as nuclear receptor co-repressors and play key roles in modulating receptor activity [26–31]. The combination of interactions among ligand, ER, other transcription factors, ERE sequences, differential recruitment of co-regulators, and the overall allosteric effects on receptor complexes allows for an intricate pattern of gene- and tissue-specific effects on target gene expression. In addition to its nuclear functions, ER has been shown to exert rapid non-genomic effects through interactions with cell membrane-associated growth factor receptors and components of downstream signal transduction pathways in the cytoplasm [32]. Post-translational modifications of ER provide additional regulatory mechanisms and enable integration of signals from multiple pathways with oestrogen signalling [33].

Adding to the mechanistic complexity and refinement of oestrogen signalling, a second *ER* gene was discovered in 1996 by Gustafsson and Kuiper and was named ERβ [34]. The original ER was renamed ERα. ERα and ERβ share a 56% similarity in their ligand-binding domains, and both bind the predominant endogenous oestrogen 17β-estradiol. The differences in their ligand-binding domains, however, also result in

selective binding of natural and synthetic ligands and allow for selective targeting of each receptor subtype. The two receptors have nearly identical DNA binding domains and share affinity for the canonical ERE. Studies of ER α -positive MCF7 breast cancer cells engineered to express ER β have confirmed a substantial overlap of DNA-binding sites between the two receptors [35–37]. Intriguingly, their similarities in DNA binding resulted in different gene expression profiles with only a minority of ER β -regulated genes also regulated by ER α [36, 38–41]. These functional differences may be due to the low conservation of their respective N-terminal AF-1 domains and their different abilities to interact with co-regulators [42]. When co-expressed, ER α and ER β can function as both homodimers and heterodimers; these complexes appear to have their own transcriptional activities and regulate distinct gene sets [43, 44].

While both receptors are found in the normal breast, ER β expression appears to be more widespread in mammary tissues [45, 46]. In both the rodent mammary gland and in the human normal breast, ER β is found in epithelial and stromal cells, while ER α is only expressed in a subset of epithelial cells [46–48]. Nonetheless, ER α is the main mediator of the oestrogen-regulated ductal elongation and growth at puberty and during the menstrual cycle, although this is at least partly a systemic effect through the hypothalamic/pituitary axis [49, 50]. ER β knockout mice have normal ductal and alveolar development [51], but ER β is involved in the final terminal differentiation of the mammary gland [47].

ER α is upregulated in the majority of breast cancers, and its expression is a hallmark of hormone-dependent tumour growth. ER β levels, in contrast, are decreased in tumour cells [52–57]. Whereas ER α is clearly linked to prognosis and response to endocrine therapy, there is no clear evidence that ER β expression is linked to clinical parameters in breast cancer. This may be due to difficulties in accurately quantifying ER β protein levels using existing reagents and techniques [58]. While oestrogen treatment of ER α -positive breast cancer cells stimulates proliferation, exogenously introduced ER β in some studies suppresses ER α -induced proliferation and transcriptional activity while also inducing independent transcriptional and functional changes [40, 41, 59–62]. Related to these anti-proliferative effects, it has also been reported that ER β -positive tumours may respond more favourably to tamoxifen, and ER β agonist treatment of ER α -positive breast cancer cell lines appear to enhance their sensitivity to tamoxifen [63, 64]. Re-introduction of ER β in more invasive ER α -negative breast cancers can, however, increase cell proliferation [65, 66]. The body of data correlating ER β to both anti-proliferative and proliferative parameters suggests a bifurcated role for ER β breast cancer biology, but the exact function of ER β in tumourigenesis and disease progression remains to be determined [66].

Targeting ER and oestrogen signalling in breast cancer prevention and treatment

For several decades following Beatson's initial published report, castration by surgical means or by irradiation was used to treat premenopausal women with recurrent or distant metastatic breast cancer. In some postmenopausal women, high doses of androgen or, paradoxically, the synthetic non-steroidal oestrogen diethylstilbestrol was effective in the treatment of advanced diseases [67–69]. Identification of ER α and the development of methodology to detect its expression by hormone binding assays in tumour samples enabled the clinical studies required that ER α be established as a prognostic marker for response to hormone therapy, and determining the ER α -status of tumour samples is now standard practice in clinical oncology [7].

The major breakthrough in targeting oestrogen signalling and ER α came from the development of non-steroidal anti-oestrogens using derivatives of triphenylethylenes by the pharmaceutical industry. The goal of these efforts was to develop anti-oestrogenic compounds that can be used in contraception. One compound, ICI 46, 474, had modest effects on fertility but showed promise as an anti-cancer agent with comparable effects with castration or hormone therapy [70]. This compound, later named tamoxifen, was shown to bind ER α , disrupt the binding of oestrogen, and block hormone-dependent breast cancer cell proliferation and tumour formation [71–73]. Following extensive pre-clinical and clinical studies, tamoxifen was approved for the treatment of ER α -positive breast cancers and for the prevention of breast cancer in high-risk individuals.

An early concern regarding the application of anti-oestrogens is their potential impact on the beneficial effects of oestrogen on bone density and cardioprotection. Interestingly, while blocking the effects of oestrogen in breast cancer cells, tamoxifen treatment actually improved bone density and reduced circulating levels of the harmful low-density lipoproteins. One of the negative effects of this selective action is that tamoxifen increases endometrial cell proliferation and risk for endometrial cancers [74]. Another non-steroidal anti-oestrogen candidate,

keoxifene, later renamed raloxifene, was demonstrated to be effective in treating osteoporosis and was also approved for the prevention of breast cancer. Compared with tamoxifen, raloxifene does not have an effect on endometrial cell growth and proliferation. Tamoxifen and raloxifene are the first members of a class of drugs, termed selective oestrogen receptor modulators (SERMs). They exhibit both oestrogenic and anti-oestrogenic effects in a tissue-specific manner and raise the possibility of simultaneously targeting multiple endocrine-related diseases or conditions. An alternative approach for directly targeting ER in breast cancer treatment is through the use of pure anti-oestrogens. Fulvestrant, initially designated as ICI 182,780, is a steroidal compound with high affinity for ER α . In addition to blocking ER activity, treatment with fulvestrant also leads to the rapid degradation of ER proteins. Consequently, treatment completely disrupts ER activity, as compared with the SERMs. This drug is particularly effective as second-line treatment when tumour cells develop resistance to tamoxifen but still require ER for continuing proliferation [75].

As the role of oestrogen became apparent in hormonal carcinogenesis and disease progression in the majority of breast cancers, an alternative strategy for targeting oestrogen signalling and ER functions emerged. Aromatase is a key enzyme involved in the conversion of androgen to oestrogen by catalysing the aromatisation of the A ring in testosterone. Inhibition of aromatase activity indirectly targets ER functions by effectively starving hormone-dependent tumour cells of locally produced oestrogens. Steroidal (exemestane) and non-steroidal (anastrozole, letrozole) aromatase inhibitors (AIs) have been developed to selectively target aromatase enzymes. These compounds either bind and inactivate aromatase or compete with endogenous substrates to reduce oestrogen production. In clinical trials, AIs showed improved efficacy as compared with treatments with tamoxifen, and these drugs are now approved for use in the adjuvant therapy of postmenopausal patients with ER-positive tumours [76–78].

Challenges and opportunities

ER α protein level, as noted previously, is the major marker for potential response to endocrine therapy. Progesterone receptor (PR), an ER α target gene, expression is an additional marker for responsiveness. Not all tumours that are classified as ER α -positive, however, respond to treatments. Resistance to endocrine therapy is estimated at about 40% [79]. The evolutionary history and specific somatic mutations that gave rise to the primary tumours may have rendered them non-responsive prior to diagnosis and subsequent treatment. Moreover, the selective pressures of long-term endocrine treatment may drive the evolution of resistant tumour cells and recurrent tumours. Mechanisms of resistance to endocrine therapy include hypersensitivity to low levels of oestrogen following treatments with AIs, alternative activation of ER α via growth factor-mediated pathways and mechanisms, and complete oestrogen- and ER α -independent growth and proliferation of tumour cells [80]. Another challenge in the application of endocrine therapy is the treatment of premenopausal patients where disruption of hormone production and ER functions may be less effective and desirable and also introduces side effects, which may increase susceptibility to other diseases following long-term treatments [81]. In spite of the benefits of current endocrine therapeutic options, further scientific and technical breakthroughs are required to fully realise the potential of targeting endocrine-related mechanisms and reducing the morbidity and mortality associated with hormone-dependent breast cancers.

Advances in genomics and genomic technologies have contributed significantly to biomedical research in general and provided a number of mechanistic insights into ER biology in breast cancer cells. These insights have resulted in candidate markers and target mechanisms in endocrine therapy. For example, gene expression profiling studies using microarrays have identified hundreds of oestrogen responsive genes, both transcriptional targets as well as those downstream of ER-regulated signalling pathways, which can be exploited as both markers of oestrogen responsiveness in tumour cells and as targetable genes and gene networks, which specifically regulate tumour cell proliferation [82–84]. Comparative analysis of sensitive and resistant cells may further elucidate markers and mechanisms of resistance. Similar gene expression studies in clinical samples have identified gene sets and signatures that define clinical subtypes and predict response to endocrine therapy and may also suggest potential resistant mechanisms [85]. Genome-wide mapping studies of ER binding sites and computational modelling of sequence motifs have identified co-localising transcription factors such as FOXA1, GATA3, and AP-2 γ that are required for ER transcriptional regulatory activity and represent additional candidate markers and therapeutic targets [13, 86, 87]. Improvements and innovations in proteomic technologies also contribute to our understanding of the ER complex, including associated co-regulators and transcription factors and may define potential markers and targets [88].

Genomic studies have also highlighted the emerging importance of non-coding RNAs in basic and translational research. Small microRNAs (miRNAs) serve as key regulators of gene expression by targeting genes for degradation or by blocking their translation. ER α -positive breast cancers display a distinct miRNA-expression profile compared with ER α -negative breast cancers [89–92]. Whether ER α directly regulates miRNA is not clear, but miRNA regulations are nonetheless likely to occur indirectly via other oestrogen-responsive genes or through ER α interaction with the miRNA processing machinery [93–95]. In addition, several miRNAs, including miR-206, have been shown to regulate ER α expression by targeting the 3' untranslated region of its mRNA [96, 97]. Transcriptome-wide nuclear run-on studies have identified long non-coding RNAs (lncRNAs) as early targets of activated ER [98]. These transcripts share the same features as protein-coding RNAs such as capping, splice sites, and polyadenylation but encode extremely short open-reading frames. Functionally, lncRNAs participate in RNA–protein, RNA–RNA, and RNA–DNA interactions in molecular processes, including those that are involved in cancer-related functions [99, 100]. Recent report by Li and colleagues shows that a specific type of lncRNAs transcribed from enhancer regions of ER target genes and named enhancer RNAs, function in the looping of chromatin that facilitates interactions between distal regulatory sites with promoters of target genes [101]. Non-coding RNAs can be specifically targeted by complementary RNAs, and their expression and function disrupted by the cellular RNA interference mechanisms [102]. The rapid progress in understanding the roles of RNAs in oestrogen signalling and ER functions suggests the potential of applying RNA therapeutics, singly or in combination with existing chemo- and endocrine therapy drugs, in improving the specificity and efficacy of endocrine therapy in breast cancer prevention and treatment. Mechanisms of oestrogen signalling and ER action and potential markers and targets are summarised in Figure 1.

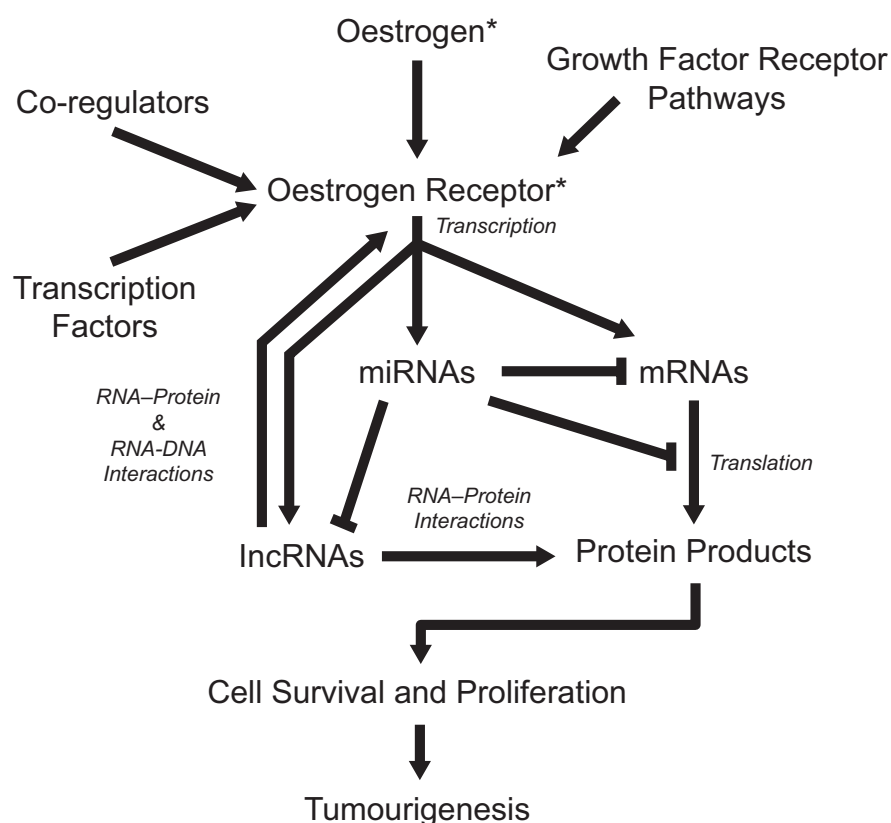


Figure 1. Summary of molecular interactions and mechanisms involved in oestrogen signalling and oestrogen receptor functions. Each component represents potential markers and target mechanisms for endocrine therapy. *Targets of current endocrine therapeutics.

Review

Conclusion

Current successes in the treatment of hormone-dependent breast cancers still leave room for significant improvements in the specificity and efficacy of current endocrine therapeutic approaches and in overcoming resistant tumours. Accumulating insights regarding oestrogen signalling and mechanisms of action of ligands and ER provide opportunities for the development of novel markers, targets, and therapeutic strategies.

Conflict of interest statement

The authors declare that they have no conflict of interest.

References

1. Jensen E (2012) **A conversation with Elwood Jensen. Interview by David D. Moore** *Annu Rev Physiol* **74** 1–11 DOI: [10.1146/annurev-physiol-020911-153327](https://doi.org/10.1146/annurev-physiol-020911-153327) PMID: [21888507](https://pubmed.ncbi.nlm.nih.gov/21888507/)
2. Beatson GT (1896) **On the treatment of inoperable cases of carcinoma of the mamma: suggestions for a new method of treatment, with illustrative cases** *Lancet* **2** 104–7 DOI: [10.1016/S0140-6736\(01\)72307-0](https://doi.org/10.1016/S0140-6736(01)72307-0)
3. Nielsen DL, Andersson M and Kamby C (2009) **HER2-targeted therapy in breast cancer. Monoclonal antibodies and tyrosine kinase inhibitors** *Cancer Treat Rev* **35**(2) 121–36 DOI: [10.1016/j.ctrv.2008.09.003](https://doi.org/10.1016/j.ctrv.2008.09.003) PMID: [19008049](https://pubmed.ncbi.nlm.nih.gov/19008049/)
4. Jensen EV and Jacobson HI (1960) **Fate of steroid estrogens in target tissues** *Biological Activities of Steroids in Relation to Cancer*, ed G Pincus, EP Vollmer (New York: Academic Press) pp 161–74
5. Jensen EV, Suzuki T, Kawashima T, Stumpf WE, Jungblut PW and DeSombre ER (1968) **A two-step mechanism for the interaction of estradiol with rat uterus** *Proc Natl Acad Sci USA* **59**(2) 632–8 DOI: [10.1073/pnas.59.2.632](https://doi.org/10.1073/pnas.59.2.632) PMID: [5238991](https://pubmed.ncbi.nlm.nih.gov/5238991/) PMCID: [224719](https://pubmed.ncbi.nlm.nih.gov/224719/)
6. Toft D and Gorski J (1966) **A receptor molecule for estrogens: isolation from the rat uterus and preliminary characterization** *Proc Natl Acad Sci USA* **55**(6) 1574–81 DOI: [10.1073/pnas.55.6.1574](https://doi.org/10.1073/pnas.55.6.1574) PMID: [5227676](https://pubmed.ncbi.nlm.nih.gov/5227676/) PMCID: [224361](https://pubmed.ncbi.nlm.nih.gov/224361/)
7. Toft D, Shyamala G and Gorski J (1967) **A receptor molecule for estrogens: studies using a cell-free system** *Proc Natl Acad Sci USA* **57**(6) 1740–3 DOI: [10.1073/pnas.57.6.1740](https://doi.org/10.1073/pnas.57.6.1740) PMID: [5232110](https://pubmed.ncbi.nlm.nih.gov/5232110/) PMCID: [224541](https://pubmed.ncbi.nlm.nih.gov/224541/)
8. O'Malley BW and Means AR (1974) **Female steroid hormones and target cell nuclei** *Science* **183**(4125) 610–20 DOI: [10.1126/science.183.4125.610](https://doi.org/10.1126/science.183.4125.610) PMID: [4359082](https://pubmed.ncbi.nlm.nih.gov/4359082/)
9. Green S, Walter P, Kumar V, Krust A, Bornert JM, Argos P, *et al* (1986) **Human oestrogen receptor cDNA: sequence, expression and homology to v-erb-A** *Nature* **320**(6058) 134–9 DOI: [10.1038/320134a0](https://doi.org/10.1038/320134a0) PMID: [3754034](https://pubmed.ncbi.nlm.nih.gov/3754034/)
10. Kumar V, Green S, Staub A and Chambon P (1986) **Localisation of the oestradiol-binding and putative DNA-binding domains of the human oestrogen receptor** *Embo J* **5**(9) 2231–6 PMID: [3780678](https://pubmed.ncbi.nlm.nih.gov/3780678/) PMCID: [1167105](https://pubmed.ncbi.nlm.nih.gov/1167105/)
11. Warnmark A, Treuter E, Wright AP and Gustafsson JA (2003) **Activation functions 1 and 2 of nuclear receptors: molecular strategies for transcriptional activation** *Mol Endocrinol* **17**(10) 1901–9 DOI: [10.1210/me.2002-0384](https://doi.org/10.1210/me.2002-0384) PMID: [12893880](https://pubmed.ncbi.nlm.nih.gov/12893880/)
12. Nilsson S and Gustafsson JA (2002) **Estrogen receptor action** *Crit Rev Eukaryot Gene Expr* **12**(4) 237–57 DOI: [10.1615/CritRevEukaryotGeneExpr.v12.i4.10](https://doi.org/10.1615/CritRevEukaryotGeneExpr.v12.i4.10) PMID: [12641394](https://pubmed.ncbi.nlm.nih.gov/12641394/)

13. Carroll JS, Liu XS, Brodsky AS, Li W, Meyer CA, Szary AJ, *et al* (2005) **Chromosome-wide mapping of estrogen receptor binding reveals long-range regulation requiring the forkhead protein FoxA1** *Cell* **122**(1) 33–43 DOI: [10.1016/j.cell.2005.05.008](https://doi.org/10.1016/j.cell.2005.05.008) PMID: [16009131](https://pubmed.ncbi.nlm.nih.gov/16009131/)
14. Shang Y, Hu X, DiRenzo J, Lazar MA and Brown M (2000) **Cofactor dynamics and sufficiency in estrogen receptor-regulated transcription** *Cell* **103**(6) 843–52 DOI: [10.1016/S0092-8674\(00\)00188-4](https://doi.org/10.1016/S0092-8674(00)00188-4) PMID: [11136970](https://pubmed.ncbi.nlm.nih.gov/11136970/)
15. Metivier R, Penot G, Hubner MR, Reid G, Brand H, Kos M, *et al* (2003) **Estrogen receptor-alpha directs ordered, cyclical, and combinatorial recruitment of cofactors on a natural target promoter** *Cell* **115**(6) 751–63 DOI: [10.1016/S0092-8674\(03\)00934-6](https://doi.org/10.1016/S0092-8674(03)00934-6) PMID: [14675539](https://pubmed.ncbi.nlm.nih.gov/14675539/)
16. Anzick SL, Kononen J, Walker RL, Azorsa DO, Tanner MM, Guan XY, *et al* (1997) **AIB1, a steroid receptor coactivator amplified in breast and ovarian cancer** *Science* **277**(5328) 965–8 DOI: [10.1126/science.277.5328.965](https://doi.org/10.1126/science.277.5328.965) PMID: [9252329](https://pubmed.ncbi.nlm.nih.gov/9252329/)
17. Phelan ML, Sif S, Narlikar GJ and Kingston RE (1999) **Reconstitution of a core chromatin remodeling complex from SWI/SNF subunits** *Mol Cell* **3**(2) 247–53 DOI: [10.1016/S1097-2765\(00\)80315-9](https://doi.org/10.1016/S1097-2765(00)80315-9) PMID: [10078207](https://pubmed.ncbi.nlm.nih.gov/10078207/)
18. Wang W, Cote J, Xue Y, Zhou S, Khavari PA, Biggar SR, *et al* (1996) **Purification and biochemical heterogeneity of the mammalian SWI-SNF complex** *Embo J* **15**(19) 5370–82 PMID: [8895581](https://pubmed.ncbi.nlm.nih.gov/8895581/) PMCID: [452280](https://pubmed.ncbi.nlm.nih.gov/452280/)
19. Halachmi S, Marden E, Martin G, MacKay H, Abbondanza C and Brown M (1994) **Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription** *Science* **264**(5164) 1455–8 DOI: [10.1126/science.8197458](https://doi.org/10.1126/science.8197458) PMID: [8197458](https://pubmed.ncbi.nlm.nih.gov/8197458/)
20. Ogryzko VV, Schiltz RL, Russanova V, Howard BH and Nakatani Y (1996) **The transcriptional coactivators p300 and CBP are histone acetyltransferases** *Cell* **87**(5) 953–9 DOI: [10.1016/S0092-8674\(00\)82001-2](https://doi.org/10.1016/S0092-8674(00)82001-2) PMID: [8945521](https://pubmed.ncbi.nlm.nih.gov/8945521/)
21. Onate SA, Tsai SY, Tsai MJ and O'Malley BW (1995) **Sequence and characterization of a coactivator for the steroid hormone receptor superfamily** *Science* **270**(5240) 1354–7 DOI: [10.1126/science.270.5240.1354](https://doi.org/10.1126/science.270.5240.1354) PMID: [7481822](https://pubmed.ncbi.nlm.nih.gov/7481822/)
22. Sterner DE and Berger SL (2000) **Acetylation of histones and transcription-related factors** *Microbiol Mol Biol Rev* **64**(2) 435–59 DOI: [10.1128/MMBR.64.2.435-459.2000](https://doi.org/10.1128/MMBR.64.2.435-459.2000) PMID: [10839822](https://pubmed.ncbi.nlm.nih.gov/10839822/) PMCID: [98999](https://pubmed.ncbi.nlm.nih.gov/98999/)
23. Chen D, Ma H, Hong H, Koh SS, Huang SM, Schurter BT, *et al* (1999) **Regulation of transcription by a protein methyltransferase** *Science* **284**(5423) 2174–7 DOI: [10.1126/science.284.5423.2174](https://doi.org/10.1126/science.284.5423.2174) PMID: [10381882](https://pubmed.ncbi.nlm.nih.gov/10381882/)
24. Lin WJ, Gary JD, Yang MC, Clarke S and Herschman HR (1996) **The mammalian immediate-early TIS21 protein and the leukemia-associated BTG1 protein interact with a protein-arginine N-methyltransferase** *J Biol Chem* **271**(25) 15034–44 DOI: [10.1074/jbc.271.25.15034](https://doi.org/10.1074/jbc.271.25.15034) PMID: [8663146](https://pubmed.ncbi.nlm.nih.gov/8663146/)
25. Wang H, Huang ZQ, Xia L, Feng Q, Erdjument-Bromage H, Strahl BD, *et al* (2001) **Methylation of histone H4 at arginine 3 facilitating transcriptional activation by nuclear hormone receptor** *Science* **293**(5531) 853–7 DOI: [10.1126/science.1060781](https://doi.org/10.1126/science.1060781) PMID: [11387442](https://pubmed.ncbi.nlm.nih.gov/11387442/)
26. Treuter E, Albrechtsen T, Johansson L, Leers J and Gustafsson JA (1998) **A regulatory role for RIP140 in nuclear receptor activation** *Mol Endocrinol* **12**(6) 864–81 DOI: [10.1210/me.12.6.864](https://doi.org/10.1210/me.12.6.864) PMID: [9626662](https://pubmed.ncbi.nlm.nih.gov/9626662/)
27. Hu X and Lazar MA (1999) **The CoRNR motif controls the recruitment of corepressors by nuclear hormone receptors** *Nature* **402**(6757) 93–6 DOI: [10.1038/47069](https://doi.org/10.1038/47069) PMID: [10573424](https://pubmed.ncbi.nlm.nih.gov/10573424/)
28. Webb P, Anderson CM, Valentine C, Nguyen P, Marimuthu A, West BL, *et al* (2000) **The nuclear receptor corepressor (N-CoR) contains three isoleucine motifs (I/LXXII) that serve as receptor interaction domains (IDs)** *Mol Endocrinol* **14**(12) 1976–85 DOI: [10.1210/me.14.12.1976](https://doi.org/10.1210/me.14.12.1976) PMID: [11117528](https://pubmed.ncbi.nlm.nih.gov/11117528/)

29. Chen JD and Evans RM (1995) **A transcriptional co-repressor that interacts with nuclear hormone receptors** *Nature* **377**(6548) 454–7 DOI: [10.1038/377454a0](https://doi.org/10.1038/377454a0) PMID: [7566127](https://pubmed.ncbi.nlm.nih.gov/7566127/)
30. Fernandes I, Bastien Y, Wai T, Nygard K, Lin R, Cormier O, *et al* (2003) **Ligand-dependent nuclear receptor corepressor LCoR functions by histone deacetylase-dependent and -independent mechanisms** *Mol Cell* **11**(1) 139–50 DOI: [10.1016/S1097-2765\(03\)00014-5](https://doi.org/10.1016/S1097-2765(03)00014-5) PMID: [12535528](https://pubmed.ncbi.nlm.nih.gov/12535528/)
31. Montano MM, Ekena K, Delage-Mourroux R, Chang W, Martini P and Katzenellenbogen BS (1999) **An estrogen receptor-selective coregulator that potentiates the effectiveness of antiestrogens and represses the activity of estrogens** *Proc Natl Acad Sci USA* **96**(12) 6947–52 DOI: [10.1073/pnas.96.12.6947](https://doi.org/10.1073/pnas.96.12.6947) PMID: [10359819](https://pubmed.ncbi.nlm.nih.gov/10359819/) PMCID: [22022](https://pubmed.ncbi.nlm.nih.gov/22022/)
32. Levin ER and Pietras RJ (2008) **Estrogen receptors outside the nucleus in breast cancer** *Breast Cancer Res Treat* **108**(3) 351–61 DOI: [10.1007/s10549-007-9618-4](https://doi.org/10.1007/s10549-007-9618-4) PMID: [17592774](https://pubmed.ncbi.nlm.nih.gov/17592774/)
33. Anbalagan M, Huderson B, Murphy L and Rowan BG (2012) **Post-translational modifications of nuclear receptors and human disease** *Nucl Recept Signal* **10** e001 PMID: [22438791](https://pubmed.ncbi.nlm.nih.gov/22438791/) PMCID: [3309075](https://pubmed.ncbi.nlm.nih.gov/3309075/)
34. Kuiper GG, Enmark E, Peltö-Huikko M, Nilsson S and Gustafsson JA (1996) **Cloning of a novel receptor expressed in rat prostate and ovary** *Proc Natl Acad Sci USA* **93**(12) 5925–30 DOI: [10.1073/pnas.93.12.5925](https://doi.org/10.1073/pnas.93.12.5925) PMID: [8650195](https://pubmed.ncbi.nlm.nih.gov/8650195/) PMCID: [39164](https://pubmed.ncbi.nlm.nih.gov/39164/)
35. Zhao C, Gao H, Liu Y, Papoutsis Z, Jaffrey S, Gustafsson JA, *et al* (2010) **Genome-wide mapping of estrogen receptor-beta-binding regions reveals extensive cross-talk with transcription factor activator protein-1** *Cancer Res* **70** 5174–83 DOI: [10.1158/0008-5472.CAN-09-4407](https://doi.org/10.1158/0008-5472.CAN-09-4407) PMID: [20501845](https://pubmed.ncbi.nlm.nih.gov/20501845/)
36. Grober OM, Mutarelli M, Giurato G, Ravo M, Cicatiello L, De Filippo MR, *et al* (2011) **Global analysis of estrogen receptor beta binding to breast cancer cell genome reveals an extensive interplay with estrogen receptor alpha for target gene regulation** *BMC Genomics* **12** 36 DOI: [10.1186/1471-2164-12-36](https://doi.org/10.1186/1471-2164-12-36) PMID: [21235772](https://pubmed.ncbi.nlm.nih.gov/21235772/) PMCID: [3025958](https://pubmed.ncbi.nlm.nih.gov/3025958/)
37. Charn TH, Liu ET, Chang EC, Lee YK, Katzenellenbogen JA and Katzenellenbogen BS (2010) **Genome-wide dynamics of chromatin binding of estrogen receptors alpha and beta: mutual restriction and competitive site selection** *Mol Endocrinol* **24** 47–59 DOI: [10.1210/me.2009-0252](https://doi.org/10.1210/me.2009-0252) PMCID: [2802902](https://pubmed.ncbi.nlm.nih.gov/2802902/)
38. Tee MK, Rogatsky I, Tzagarakis-Foster C, Cvoro A, An J, Christy RJ, *et al* (2004) **Estradiol and selective estrogen receptor modulators differentially regulate target genes with estrogen receptors alpha and beta** *Mol Biol Cell* **15** 1262–72 DOI: [10.1091/mbc.E03-06-0360](https://doi.org/10.1091/mbc.E03-06-0360) PMCID: [PMC363122](https://pubmed.ncbi.nlm.nih.gov/PMC363122/)
39. Stossi F, Barnett DH, Frasor J, Komm B, Lyttle CR and Katzenellenbogen BS (2004) **Transcriptional profiling of estrogen-regulated gene expression via estrogen receptor (ER) {alpha} or ER{beta} in human osteosarcoma cells: distinct and common target genes for these receptors** *Endocrinology* **145**(7) 3473–86 DOI: [10.1210/en.2003-1682](https://doi.org/10.1210/en.2003-1682) PMID: [15033914](https://pubmed.ncbi.nlm.nih.gov/15033914/)
40. Chang EC, Frasor J, Komm B and Katzenellenbogen BS (2006) **Impact of Estrogen receptor beta on gene networks regulated by estrogen receptor alpha in breast cancer cells** *Endocrinology* **147**(10) 4831–42 DOI: [10.1210/en.2006-0563](https://doi.org/10.1210/en.2006-0563) PMID: [16809442](https://pubmed.ncbi.nlm.nih.gov/16809442/)
41. Williams C, Edvardsson K, Lewandowski SA, Strom A and Gustafsson J-A (2008) **A genome-wide study of the repressive effects of estrogen receptor beta on estrogen receptor alpha signaling in breast cancer cells** *Oncogene* **27** 1019–32 DOI: [10.1038/sj.onc.1210712](https://doi.org/10.1038/sj.onc.1210712) PMID: [17700529](https://pubmed.ncbi.nlm.nih.gov/17700529/)
42. Mosselman S, Polman J and Dijkema R (1996) **ER beta: identification and characterization of a novel human estrogen receptor** *FEBS Lett* **392**(1) 49–53 DOI: [10.1016/0014-5793\(96\)00782-X](https://doi.org/10.1016/0014-5793(96)00782-X) PMID: [8769313](https://pubmed.ncbi.nlm.nih.gov/8769313/)
43. Monroe DG, Secreto FJ, Subramaniam M, Getz BJ, Khosla S and Spelsberg TC (2005) **Estrogen receptor {alpha} and {beta} heterodimers exert unique effects on estrogen- and tamoxifen-dependent gene expression in human U2OS osteosarcoma cells** *Mol Endocrinol* **19**(6) 1555–68 DOI: [10.1210/me.2004-0381](https://doi.org/10.1210/me.2004-0381) PMID: [15802376](https://pubmed.ncbi.nlm.nih.gov/15802376/)

44. Papoutsis Z, Zhao C, Putnik M, Gustafsson J and Dahlman-Wright K (2009) **Binding of estrogen receptor alpha/beta heterodimers to chromatin in MCF-7 cells** *J Mol Endocrinol* **43**(2) 65–72 DOI: [10.1677/JME-08-0177](https://doi.org/10.1677/JME-08-0177) PMID: [19376833](https://pubmed.ncbi.nlm.nih.gov/19376833/)
45. Speirs V, Skliris GP, Burdall SE and Carder PJ (2002) **Distinct expression patterns of ER alpha and ER beta in normal human mammary gland** *J Clin Pathol* **55**(5) 371–4 DOI: [10.1136/jcp.55.5.371](https://doi.org/10.1136/jcp.55.5.371) PMID: [11986344](https://pubmed.ncbi.nlm.nih.gov/11986344/) PMCID: [1769648](https://pubmed.ncbi.nlm.nih.gov/1769648/)
46. Li S, Han B, Liu G, Ouellet J, Labrie F and Pelletier G (2010) **Immunocytochemical localization of sex steroid hormone receptors in normal human mammary gland** *J Histochem Cytochem* **58** 509–15 DOI: [10.1369/jhc.2009.954644](https://doi.org/10.1369/jhc.2009.954644) PMCID: [2874183](https://pubmed.ncbi.nlm.nih.gov/2874183/)
47. Cheng G, Weihua Z, Warner M and Gustafsson J-A (2004) **Inaugural article: estrogen receptors ER{alpha} and ER{beta} in proliferation in the rodent mammary gland** *PNAS* **101**(11) 3739–46 DOI: [10.1073/pnas.0307864100](https://doi.org/10.1073/pnas.0307864100) PMID: [14762170](https://pubmed.ncbi.nlm.nih.gov/14762170/) PMCID: [374314](https://pubmed.ncbi.nlm.nih.gov/374314/)
48. Palmieri C, Saji S, Sakaguchi H, Cheng G, Sunters A, O'Hare MJ, *et al* (2004) **The expression of ERb and its variants, but not ERa, in adult human mammary fibroblasts** *J Mol Endocrinol* **33**(1) 35–50 DOI: [10.1677/jme.0.0330035](https://doi.org/10.1677/jme.0.0330035) PMID: [15291741](https://pubmed.ncbi.nlm.nih.gov/15291741/)
49. Bocchinfuso WP, Lindzey JK, Hewitt SC, Clark JA, Myers PH, Cooper R, *et al* (2000) **Induction of mammary gland development in estrogen receptor-a knockout mice** *Endocrinology* **141** 2982–94 DOI: [10.1210/en.141.8.2982](https://doi.org/10.1210/en.141.8.2982) PMID: [10919287](https://pubmed.ncbi.nlm.nih.gov/10919287/)
50. Hennighausen L and Robinson GW (2001) **Signaling pathways in mammary gland development** *Dev Cell* **1** 467–75 DOI: [10.1016/S1534-5807\(01\)00064-8](https://doi.org/10.1016/S1534-5807(01)00064-8) PMID: [11703938](https://pubmed.ncbi.nlm.nih.gov/11703938/)
51. Forster C, Makela S, Warri A, Kietz S, Becker D, Hultenby K, *et al* (2002) **Involvement of estrogen receptor beta in terminal differentiation of mammary gland epithelium** *Proc Natl Acad Sci USA* **99** 15578–83 DOI: [10.1073/pnas.192561299](https://doi.org/10.1073/pnas.192561299) PMID: [12438700](https://pubmed.ncbi.nlm.nih.gov/12438700/) PMCID: [137759](https://pubmed.ncbi.nlm.nih.gov/137759/)
52. Roger P, Sahla ME, Mäkelä S, Gustafsson JA, Baldet P and Rochefort H (2001) **Decreased expression of estrogen receptor beta protein in proliferative preinvasive mammary tumors** *Cancer Res* **61**(6) 2537–41 PMID: [11289127](https://pubmed.ncbi.nlm.nih.gov/11289127/)
53. Palmieri C, Cheng G, Saji S, Zelada-Hedman M, Warri A, Weihua Z, *et al* (2002) **Estrogen receptor beta in breast cancer** *Endocr Relat Cancer* **9**(1) 1–13 DOI: [10.1677/erc.0.0090001](https://doi.org/10.1677/erc.0.0090001) PMID: [11914179](https://pubmed.ncbi.nlm.nih.gov/11914179/)
54. Shaaban AM, O'Neill PA, Davies MP, Sibson R, West CR, Smith PH, *et al* (2003) **Declining estrogen receptor-beta expression defines malignant progression of human breast neoplasia** *Am J Surg Pathol* **27** 1502–12 DOI: [10.1097/00000478-200312000-00002](https://doi.org/10.1097/00000478-200312000-00002) PMID: [14657709](https://pubmed.ncbi.nlm.nih.gov/14657709/)
55. Platet N, Cathiard AM, Gleizes M and Garcia M (2004) **Estrogens and their receptors in breast cancer progression: a dual role in cancer proliferation and invasion** *Crit Rev Oncol Hematol* **51** 55–67 DOI: [10.1016/j.critrevonc.2004.02.001](https://doi.org/10.1016/j.critrevonc.2004.02.001) PMID: [15207254](https://pubmed.ncbi.nlm.nih.gov/15207254/)
56. Saji S, Hirose M and Toi M (2005) **Clinical significance of estrogen receptor beta in breast cancer** *Cancer Chemother Pharmacol* **56** 21–6 DOI: [10.1007/s00280-005-0107-3](https://doi.org/10.1007/s00280-005-0107-3) PMID: [16273360](https://pubmed.ncbi.nlm.nih.gov/16273360/)
57. Zhao C, Dahlman-Wright K and Gustafsson JA (2008) **Estrogen receptor beta: an overview and update** *Nucl Recept Signal* **6**(e003) PMID: [18301783](https://pubmed.ncbi.nlm.nih.gov/18301783/) PMCID: [2254331](https://pubmed.ncbi.nlm.nih.gov/2254331/)
58. Haldosen LA, Zhao C and Dahlman-Wright K (2013) **Estrogen receptor beta in breast cancer** *Mol Cell Endocrinol* DOI: [10.1016/j.mce.2013.08.005](https://doi.org/10.1016/j.mce.2013.08.005) PMID: [23954741](https://pubmed.ncbi.nlm.nih.gov/23954741/)
59. Paech K, Webb P, Kuiper GG, Nilsson S, Gustafsson J, Kushner PJ, *et al* (1997) **Differential ligand activation of estrogen receptors ERalpha and ERbeta at AP1 sites** *Science* **277** 1508–10 DOI: [10.1126/science.277.5331.1508](https://doi.org/10.1126/science.277.5331.1508) PMID: [9278514](https://pubmed.ncbi.nlm.nih.gov/9278514/)
60. Strom A, Hartman J, Foster JS, Kietz S, Wimalasena J and Gustafsson J-A (2004) **Estrogen receptor {beta} inhibits 17{beta}-estradiol-stimulated proliferation of the breast cancer cell line T47D** *PNAS* **101**(6) 1566–71 DOI: [10.1073/pnas.0308319100](https://doi.org/10.1073/pnas.0308319100)
61. Hartman J, Lindberg K, Morani A, Inzunza J, Strom A, Gustafsson J-A (2006) **Estrogen receptor {beta} inhibits angiogenesis and growth of T47D Breast cancer xenografts** *Cancer Res* **66**(23) 11207–13 DOI: [10.1158/0008-5472.CAN-06-0017](https://doi.org/10.1158/0008-5472.CAN-06-0017) PMID: [17145865](https://pubmed.ncbi.nlm.nih.gov/17145865/)

62. Matthews J, Wihlen B, Tujague M, Wan J, Strom A and Gustafsson J-A (2006) **Estrogen receptor (er) {beta} modulates er{alpha}-mediated transcriptional activation by altering the recruitment of c-Fos and c-Jun to estrogen-responsive promoters** *Mol Endocrinol* **20**(3) 534–43 DOI: [10.1210/me.2005-0140](https://doi.org/10.1210/me.2005-0140)
63. Honma N, Horii R, Iwase T, Saji S, Younes M, Takubo K, *et al* (2008) **Clinical importance of estrogen receptor-beta evaluation in breast cancer patients treated with adjuvant tamoxifen therapy** *J Clin Oncol* **26**(22) 3727–34 DOI: [10.1200/JCO.2007.14.2968](https://doi.org/10.1200/JCO.2007.14.2968) PMID: [18669459](https://pubmed.ncbi.nlm.nih.gov/18669459/)
64. Latratch C, Schuler S, Haring J, Skrzypczak M, Ortmann O and Treeck O (2013) **Effects of a combined treatment with tamoxifen and estrogen receptor beta agonists on human breast cancer cell lines** *Arch Gynecol Obstet* DOI: [10.1007/s00404-013-2977-7](https://doi.org/10.1007/s00404-013-2977-7) PMID: [23907354](https://pubmed.ncbi.nlm.nih.gov/23907354/)
65. Tonetti DA, Rubenstein R, DeLeon M, Zhao H, Pappas SG, Bentrem DJ, *et al* (2003) **Stable transfection of an estrogen receptor beta cDNA isoform into MDA-MB-231 breast cancer cells** *J Steroid Biochem Mol Biol* **87**(1) 47–55 DOI: [10.1016/j.jsbmb.2003.07.003](https://doi.org/10.1016/j.jsbmb.2003.07.003) PMID: [14630090](https://pubmed.ncbi.nlm.nih.gov/14630090/)
66. Leygue E and Murphy LC (2013) **A bi-faceted role of estrogen receptor beta in breast cancer** *Endocr Relat Cancer* **20**(3) R127–39 DOI: [10.1530/ERC-12-0389](https://doi.org/10.1530/ERC-12-0389) PMID: [23533249](https://pubmed.ncbi.nlm.nih.gov/23533249/)
67. Kennedy BJ (1965) **Hormone therapy for advanced breast cancer** *Cancer* **18**(12) 1551–7 DOI: [10.1002/1097-0142-\(196512\)18:12<1551::AID-CNCR2820181206>3.0.CO;2-1](https://doi.org/10.1002/1097-0142-(196512)18:12<1551::AID-CNCR2820181206>3.0.CO;2-1) PMID: [5845796](https://pubmed.ncbi.nlm.nih.gov/5845796/)
68. Haddow A, Watkinson JM, Paterson E and Koller PC (1944) **Influence of synthetic oestrogens on advanced malignant disease** *Br Med J* **2**(4368) 393–8 DOI: [10.1136/bmj.2.4368.393](https://doi.org/10.1136/bmj.2.4368.393) PMID: [20785660](https://pubmed.ncbi.nlm.nih.gov/20785660/) PMCID: [2286289](https://pubmed.ncbi.nlm.nih.gov/2286289/)
69. Peethambaram PP, Ingle JN, Suman VJ, Hartmann LC and Loprinzi CL (1999) **Randomized trial of diethylstilbestrol vs. tamoxifen in postmenopausal women with metastatic breast cancer. An updated analysis** *Breast Cancer Res Treat* **54**(2) 117–22 DOI: [10.1023/A:1006185805079](https://doi.org/10.1023/A:1006185805079) PMID: [10424402](https://pubmed.ncbi.nlm.nih.gov/10424402/)
70. Cole MP, Jones CT and Todd ID (1971) **A new anti-oestrogenic agent in late breast cancer. An early clinical appraisal of ICI46474** *Br J Cancer* **25**(2) 270–5 DOI: [10.1038/bjc.1971.33](https://doi.org/10.1038/bjc.1971.33) PMID: [5115829](https://pubmed.ncbi.nlm.nih.gov/5115829/) PMCID: [2008453](https://pubmed.ncbi.nlm.nih.gov/2008453/)
71. Gottardis MM, Robinson SP and Jordan VC (1988) **Estradiol-stimulated growth of MCF-7 tumors implanted in athymic mice: a model to study the tumorigenic action of tamoxifen** *J Steroid Biochem* **30**(1–6) 311–4 DOI: [10.1016/0022-4731\(88\)90113-6](https://doi.org/10.1016/0022-4731(88)90113-6) PMID: [3386259](https://pubmed.ncbi.nlm.nih.gov/3386259/)
72. Jordan VC and Koerner S (1975) **Inhibition of oestradiol binding to mouse uterine and vaginal oestrogen receptors by triphenylethylenes** *J Endocrinol* **64**(1) 193–4 DOI: [10.1677/joe.0.0640193](https://doi.org/10.1677/joe.0.0640193) PMID: [163879](https://pubmed.ncbi.nlm.nih.gov/163879/)
73. Lippman ME and Bolan G (1975) **Oestrogen-responsive human breast cancer in long term tissue culture** *Nature* **256**(5518) 592–3 DOI: [10.1038/256592a0](https://doi.org/10.1038/256592a0) PMID: [170527](https://pubmed.ncbi.nlm.nih.gov/170527/)
74. Jordan VC, Gottardis MM and Satyaswaroop PG (1991) **Tamoxifen-stimulated growth of human endometrial carcinoma** *Ann NY Acad Sci* **622** 439–46 DOI: [10.1111/j.1749-6632.1991.tb37886.x](https://doi.org/10.1111/j.1749-6632.1991.tb37886.x) PMID: [1905895](https://pubmed.ncbi.nlm.nih.gov/1905895/)
75. Osborne CK, Pippen J, Jones SE, Parker LM, Ellis M, Come S, *et al* (2002) **Double-blind, randomized trial comparing the efficacy and tolerability of fulvestrant versus anastrozole in postmenopausal women with advanced breast cancer progressing on prior endocrine therapy: results of a North American trial** *J Clin Oncol* **20**(16) 3386–95 DOI: [10.1200/JCO.2002.10.058](https://doi.org/10.1200/JCO.2002.10.058) PMID: [12177098](https://pubmed.ncbi.nlm.nih.gov/12177098/)
76. Baum M, Budzar AU, Cuzick J, Forbes J, Houghton JH, Klijn JG, *et al* (2002) **Anastrozole alone or in combination with tamoxifen versus tamoxifen alone for adjuvant treatment of postmenopausal women with early breast cancer: first results of the ATAC randomised trial** *Lancet* **359**(9324) 2131–9 DOI: [10.1016/S0140-6736\(02\)09088-8](https://doi.org/10.1016/S0140-6736(02)09088-8) PMID: [12090977](https://pubmed.ncbi.nlm.nih.gov/12090977/)

77. Coombes RC, Hall E, Gibson LJ, Paridaens R, Jassem J, Delozier T, *et al* (2004) **A randomized trial of exemestane after two to three years of tamoxifen therapy in postmenopausal women with primary breast cancer** *N Engl J Med* **350**(11) 1081–92 DOI: [10.1056/NEJMoa040331](https://doi.org/10.1056/NEJMoa040331) PMID: [15014181](https://pubmed.ncbi.nlm.nih.gov/15014181/)
78. Goss PE, Ingle JN, Martino S, Robert NJ, Muss HB, Piccart MJ, *et al* (2003) **A randomized trial of letrozole in postmenopausal women after five years of tamoxifen therapy for early-stage breast cancer** *N Engl J Med* **349**(19) 1793–802 DOI: [10.1056/NEJMoa032312](https://doi.org/10.1056/NEJMoa032312) PMID: [14551341](https://pubmed.ncbi.nlm.nih.gov/14551341/)
79. Clarke R, Liu MC, Bouker KB, Gu Z, Lee RY, Zhu Y, *et al* (2003) **Antiestrogen resistance in breast cancer and the role of estrogen receptor signaling** *Oncogene* **22**(47) 7316–39 DOI: [10.1038/sj.onc.1206937](https://doi.org/10.1038/sj.onc.1206937) PMID: [14576841](https://pubmed.ncbi.nlm.nih.gov/14576841/)
80. Normanno N, Di Maio M, De Maio E, De Luca A, de Matteis A, Giordano A, *et al* (2005) **Mechanisms of endocrine resistance and novel therapeutic strategies in breast cancer** *Endocr Relat Cancer* **12**(4) 721–47 DOI: [10.1677/erc.1.00857](https://doi.org/10.1677/erc.1.00857) PMID: [16322319](https://pubmed.ncbi.nlm.nih.gov/16322319/)
81. Emens LA and Davidson NE (2003) **Adjuvant hormonal therapy for premenopausal women with breast cancer** *Clin Cancer Res* **9**(1 Pt 2) 486S–94S PMID: [12538505](https://pubmed.ncbi.nlm.nih.gov/12538505/)
82. Frasor J, Danes JM, Komm B, Chang KC, Lyttle CR and Katzenellenbogen BS (2003) **Profiling of estrogen up- and down-regulated gene expression in human breast cancer cells: insights into gene networks and pathways underlying estrogenic control of proliferation and cell phenotype** *Endocrinology* **144**(10) 4562–74 DOI: [10.1210/en.2003-0567](https://doi.org/10.1210/en.2003-0567) PMID: [12959972](https://pubmed.ncbi.nlm.nih.gov/12959972/)
83. Lin CY, Strom A, Vega VB, Kong SL, Yeo AL, Thomsen JS, *et al* (2004) **Discovery of estrogen receptor alpha target genes and response elements in breast tumor cells** *Genome Biol* **5**(9) R66 DOI: [10.1186/gb-2004-5-9-r66](https://doi.org/10.1186/gb-2004-5-9-r66) PMID: [15345050](https://pubmed.ncbi.nlm.nih.gov/15345050/) PMCID: [522873](https://pubmed.ncbi.nlm.nih.gov/522873/)
84. Alvarez-Baron CP, Jonsson P, Thomas C, Dryer SE and Williams C (2011) **The two-pore domain potassium channel KCNK5: induction by estrogen receptor alpha and role in proliferation of breast cancer cells** *Mol Endocrinol* **25**(8) 1326–36 DOI: [10.1210/me.2011-0045](https://doi.org/10.1210/me.2011-0045) PMID: [21680658](https://pubmed.ncbi.nlm.nih.gov/21680658/) PMCID: [3146246](https://pubmed.ncbi.nlm.nih.gov/3146246/)
85. Loi S, Piccart M and Sotiriou C (2007) **The use of gene-expression profiling to better understand the clinical heterogeneity of estrogen receptor positive breast cancers and tamoxifen response** *Crit Rev Oncol Hematol* **61**(3) 187–94 DOI: [10.1016/j.critrevonc.2006.09.005](https://doi.org/10.1016/j.critrevonc.2006.09.005) PMID: [17088071](https://pubmed.ncbi.nlm.nih.gov/17088071/)
86. Kong SL, Li G, Loh SL, Sung WK and Liu ET (2011) **Cellular reprogramming by the conjoint action of ERalpha, FOXA1, and GATA3 to a ligand-inducible growth state** *Mol Syst Biol* **7** 526 DOI: [10.1038/msb.2011.59](https://doi.org/10.1038/msb.2011.59) PMID: [21878914](https://pubmed.ncbi.nlm.nih.gov/21878914/) PMCID: [3202798](https://pubmed.ncbi.nlm.nih.gov/3202798/)
87. Tan SK, Lin ZH, Chang CW, Varang V, Chng KR, Pan YF, *et al* (2011) **AP-2gamma regulates oestrogen receptor-mediated long-range chromatin interaction and gene transcription** *Embo J* **30**(13) 2569–81 DOI: [10.1038/emboj.2011.151](https://doi.org/10.1038/emboj.2011.151) PMID: [21572391](https://pubmed.ncbi.nlm.nih.gov/21572391/) PMCID: [3155293](https://pubmed.ncbi.nlm.nih.gov/3155293/)
88. Mohammed H, D'Santos C, Serandour AA, Ali HR, Brown GD, Atkins A, *et al* (2013) **Endogenous purification reveals GREB1 as a key estrogen receptor regulatory factor** *Cell Rep* **3**(2) 342–9 DOI: [10.1016/j.celrep.2013.01.010](https://doi.org/10.1016/j.celrep.2013.01.010) PMID: [23403292](https://pubmed.ncbi.nlm.nih.gov/23403292/)
89. Bhat-Nakshatri P, Wang G, Collins NR, Thomson MJ, Geistlinger TR, Carroll JS, *et al* (2009) **Estradiol-regulated microRNAs control estradiol response in breast cancer cells** *Nucleic Acids Res* **37**(14) 4850–61 DOI: [10.1093/nar/gkp500](https://doi.org/10.1093/nar/gkp500) PMID: [19528081](https://pubmed.ncbi.nlm.nih.gov/19528081/) PMCID: [2724297](https://pubmed.ncbi.nlm.nih.gov/2724297/)
90. Blenkiron C, Goldstein LD, Thorne NP, Spiteri I, Chin SF, Dunning MJ, *et al* (2007) **MicroRNA expression profiling of human breast cancer identifies new markers of tumor subtype** *Genome Biol* **8**(10) R214 DOI: [10.1186/gb-2007-8-10-r214](https://doi.org/10.1186/gb-2007-8-10-r214) PMID: [17922911](https://pubmed.ncbi.nlm.nih.gov/17922911/) PMCID: [2246288](https://pubmed.ncbi.nlm.nih.gov/2246288/)

91. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, *et al* (2005) **MicroRNA gene expression deregulation in human breast cancer** *Cancer Res* **65**(16) 7065–70 DOI: [10.1158/0008-5472.CAN-05-1783](https://doi.org/10.1158/0008-5472.CAN-05-1783) PMID: [16103053](https://pubmed.ncbi.nlm.nih.gov/16103053/)
92. Mattie MD, Benz CC, Bowers J, Sensinger K, Wong L, Scott GK, *et al* (2006) **Optimized high-throughput microRNA expression profiling provides novel biomarker assessment of clinical prostate and breast cancer biopsies** *Mol Cancer* **5** 24 DOI: [10.1186/1476-4598-5-24](https://doi.org/10.1186/1476-4598-5-24) PMID: [16784538](https://pubmed.ncbi.nlm.nih.gov/16784538/) PMCID: [1563474](https://pubmed.ncbi.nlm.nih.gov/1563474/)
93. Katchy A, Edvardsson K, Aydogdu E, Williams C (2012) **Estradiol-activated estrogen receptor alpha does not regulate mature microRNAs in T47D breast cancer cells** *J Steroid Biochem Mol Biol* **128**(3–5) 145–53 DOI: [10.1016/j.jsbmb.2011.10.008](https://doi.org/10.1016/j.jsbmb.2011.10.008) PMID: [22079223](https://pubmed.ncbi.nlm.nih.gov/22079223/)
94. Klinge CM (2012) **miRNAs and estrogen action** *Trends Endocrinol Metab* **23**(5) 223–33 DOI: [10.1016/j.tem.2012.03.002](https://doi.org/10.1016/j.tem.2012.03.002) PMID: [22503553](https://pubmed.ncbi.nlm.nih.gov/22503553/) PMCID: [3348384](https://pubmed.ncbi.nlm.nih.gov/3348384/)
95. Yamagata K, Fujiyama S, Ito S, Ueda T, Murata T, Naitou M, *et al* (2009) **Maturation of microRNA is hormonally regulated by a nuclear receptor** *Mol Cell* **36**(2) 340–7 DOI: [10.1016/j.molcel.2009.08.017](https://doi.org/10.1016/j.molcel.2009.08.017) PMID: [19854141](https://pubmed.ncbi.nlm.nih.gov/19854141/)
96. Adams BD, Furneaux H, White BA (2007) **The micro-ribonucleic acid (miRNA) miR-206 targets the human estrogen receptor-alpha (ERalpha) and represses ERalpha messenger RNA and protein expression in breast cancer cell lines** *Mol Endocrinol* **21**(5) 1132–47 DOI: [10.1210/me.2007-0022](https://doi.org/10.1210/me.2007-0022) PMID: [17312270](https://pubmed.ncbi.nlm.nih.gov/17312270/)
97. Zhao JJ, Lin J, Yang H, Kong W, He L, Ma X, *et al* (2008) **MicroRNA-221/222 negatively regulates estrogen receptor alpha and is associated with tamoxifen resistance in breast cancer** *J Biol Chem* **283**(45) 31079–86 DOI: [10.1074/jbc.M806041200](https://doi.org/10.1074/jbc.M806041200) PMID: [18790736](https://pubmed.ncbi.nlm.nih.gov/18790736/) PMCID: [2576549](https://pubmed.ncbi.nlm.nih.gov/2576549/)
98. Hah N, Danko CG, Core L, Waterfall JJ, Siepel A, Lis JT, *et al* (2011) **A rapid, extensive, and transient transcriptional response to estrogen signaling in breast cancer cells** *Cell* **145**(4) 622–34 DOI: [10.1016/j.cell.2011.03.042](https://doi.org/10.1016/j.cell.2011.03.042) PMID: [21549415](https://pubmed.ncbi.nlm.nih.gov/21549415/) PMCID: [3099127](https://pubmed.ncbi.nlm.nih.gov/3099127/)
99. Ponting CP, Oliver PL and Reik W (2009) **Evolution and functions of long noncoding RNAs** *Cell* **136**(4) 629–41 DOI: [10.1016/j.cell.2009.02.006](https://doi.org/10.1016/j.cell.2009.02.006) PMID: [19239885](https://pubmed.ncbi.nlm.nih.gov/19239885/)
100. Lipovich L, Johnson R and Lin CY (2010) **MacroRNA underdogs in a microRNA world: evolutionary, regulatory, and biomedical significance of mammalian long non-protein-coding RNA** *Biochim Biophys Acta* **1799**(9) 597–615 DOI: [10.1016/j.bbagr.2010.10.001](https://doi.org/10.1016/j.bbagr.2010.10.001) PMID: [20951849](https://pubmed.ncbi.nlm.nih.gov/20951849/)
101. Li W, Notani D, Ma Q, Tanasa B, Nunez E, Chen AY, *et al* (2013) **Functional roles of enhancer RNAs for oestrogen-dependent transcriptional activation** *Nature* **498**(7455) 516–20 DOI: [10.1038/nature12210](https://doi.org/10.1038/nature12210) PMID: [23728302](https://pubmed.ncbi.nlm.nih.gov/23728302/) PMCID: [3718886](https://pubmed.ncbi.nlm.nih.gov/3718886/)
102. Prensner JR and Chinnaiyan AM (2011) **The emergence of lncRNAs in cancer biology** *Cancer Discov* **1**(5) 391–407 DOI: [10.1158/2159-8290.CD-11-0209](https://doi.org/10.1158/2159-8290.CD-11-0209) PMID: [22096659](https://pubmed.ncbi.nlm.nih.gov/22096659/) PMCID: [3215093](https://pubmed.ncbi.nlm.nih.gov/3215093/)