

COVID-19 vaccination immune response in patients with solid organ and haematologic malignancies: call for active monitoring

Laudy Chehade¹, Jad Zeitoun¹, Rachelle Bejjany¹, Maya Charafeddine¹, Firas Kreidieh¹, Mona Hassan¹, Ali Taher¹, Nagi El Saghir¹, Ali Shamseddine¹, Ziad Salem¹, Sally Temraz¹, Arafat Tfayli¹, Hazem Assi¹, Ali Bazarbachi¹, Jean El Cheikh¹, Iman Abou Dalle¹, Nesrine Rizk², Rami Mahfouz³ and Deborah Mukherji¹

¹Naef K. Basile Cancer Institute, American University of Beirut Medical Center, PO Box 11-0236, Riad El Solh, Beirut 1107 2020, Lebanon

²Division of Infectious Diseases, Department of Internal Medicine, American University of Beirut Medical Center, PO Box 11-0236, Riad El Solh, Beirut 1107 2020, Lebanon

³Department of Pathology and Laboratory Medicine, American University of Beirut Medical Center, PO Box 11-0236, Riad El Solh, Beirut 1107 2020, Lebanon

Abstract

Vaccines against COVID-19 have demonstrated a remarkable efficacy in decreasing hospitalisations and deaths; however, clinical trials leading to vaccine approvals did not include immunocompromised individuals such as patients receiving antineoplastic therapies. Emerging data suggest that patients on active anti-cancer therapy may have a reduced immune response to COVID-19 vaccination compared to the general population and may be at greater risk of COVID-19 infection as measures to reduce transmission in the community are relaxed. We report preliminary data from the American University of Beirut Medical Center in Lebanon demonstrating relatively low seroconversion rates. Of 36 patients on active anti-cancer therapy who had received two doses of vaccine, 17% were negative for Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) anti-spike IgG. These results highlight the importance of maintaining strict precautionary measures against COVID-19 in patients on immunosuppressive treatment. There is an urgent need for active monitoring of immune response post-vaccination in prospective studies involving populations from diverse resource settings.

Keywords: COVID-19, vaccine, chemotherapy, immune response

Despite the lack of data on the efficacy of COVID-19 vaccination in patients on anti-neoplastic therapy from registration trials, vaccination in this vulnerable population is strongly recommended due to the increased risk of severe COVID-19 infection [1].

Data from case series in high-income countries suggest that the proportion of patients with cancer on active therapy who do not develop antibodies after two doses of the Pfizer BNT162b2 mRNA COVID-19 vaccine ranges from 6% to 10% [2, 3]. To date, we have limited data on the duration of immune response in patients on active anti-cancer therapy or the potential benefit of vaccine booster doses in patients with an insufficient or short-lived immune response. Ehmsen *et al* [4] recently reported a three-fold reduction approximately in antibody titers after a period of 12 weeks post vaccination compared

Correspondence to: Deborah Mukherji
Email: dm25@aub.edu.lb

ecancer 2021, 15:1284
<https://doi.org/10.3332/ecancer.2021.1284>

Published: 07/09/2021

Received: 26/07/2021

Publication costs for this article were supported by ecancer (UK Charity number 1176307).

Copyright: © the authors; licensee ecancermedicalscience. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

to 5 weeks post vaccination; however, more data are needed to support this finding. As COVID-19 infection control measures in the community are relaxed, there is a concern that immunosuppressed patients remain at increased risk of COVID-19 infection despite being offered vaccination.

The COVID-19 and Cancer Taskforce has previously recommended a global vaccine-response monitoring programme and a draft protocol has been published [5]. We report preliminary data from the American University of Beirut Medical Center in Lebanon, where the economic crisis and currency depreciation have made access to essential medications and other resources to support the healthcare system increasingly limited.

Following institutional review board approval of the protocol, patients diagnosed with a solid organ or haematological malignancy who were on active systemic treatment at the time of planned vaccination (chemotherapy, targeted therapy, immune checkpoint inhibitor therapy, endocrine therapy) at the American University of Beirut Medical Center, Beirut, Lebanon, were included. Following written informed consent, patients were tested for COVID-19 immunoglobulin G (IgG) using a chemiluminescent microparticle immunoassay developed by Abbot Diagnostics, which measures IgG antibodies against the spike receptor-binding domain of SARS-CoV-2 (cutoff for positive test is 50 AU/mL). This assay has a specificity of 99.6% and a sensitivity of 99.35% [6].

Fifty patients were recruited with a median age of 64.5 years, 48% were males and 52% were females. Seven patients (14%) had COVID-19 infection prior to vaccination. 12% (n = 6) of the patients had a haematological malignancy and the rest had a solid organ tumour, with the following distribution: gastrointestinal 42%, breast 24%, lung 16% and genitourinary 6% (Table 1). All patients have received at least one dose of the BNT162b2 mRNA vaccine except one patient who received ChAdOx1 (two doses), one patient who received Gam-COVID-Vac (two doses) and one patient who received both BBIBP-CorV and BNT162b2 vaccines. Six patients (12%) had negative IgG titers after receiving two doses of vaccine.

Table 1. Clinical characteristics of the study cohort.

N	50
Age, years, median	64.5
Gender	
Male	24 (48%)
Female	26 (52%)
Type of malignancy	
Breast	12 (24%)
Lung	8 (16%)
Gastrointestinal	21 (42%)
Genitourinary	3 (6%)
Haematologic	6 (12%)
Types of treatment	
Chemotherapy	20 (40%)
Immunotherapy	6 (12%)
Chemotherapy + immunotherapy	5 (10%)
Chemotherapy + anti VEGF	5 (10%)
Anti CD20	2 (4%)
Anti CD38 + proteasome inhibitor	2 (4%)
Anti HER-2	3 (6%)

Table 1. Clinical characteristics of the study cohort. (Continued)

Chemotherapy + anti HER-2	2 (4%)
Endocrine + CDK4/6 inhibitor	1 (2%)
Anti EGFR	1 (2%)
Chemotherapy + anti EGFR	2 (4%)
Bispecific T cell engager	1 (2%)
Pre-vaccine COVID infection	
No	43 (86%)
Yes	7 (14%)
Vaccine information	
One dose	14 (28%)
Two doses	36 (72%)
Days between last dose and IgG measurement, median	20

VEGF: vascular endothelial growth factor; HER: human epidermal growth factor receptor; CDK: cyclin-dependent kinase; EGFR: epidermal growth factor receptor

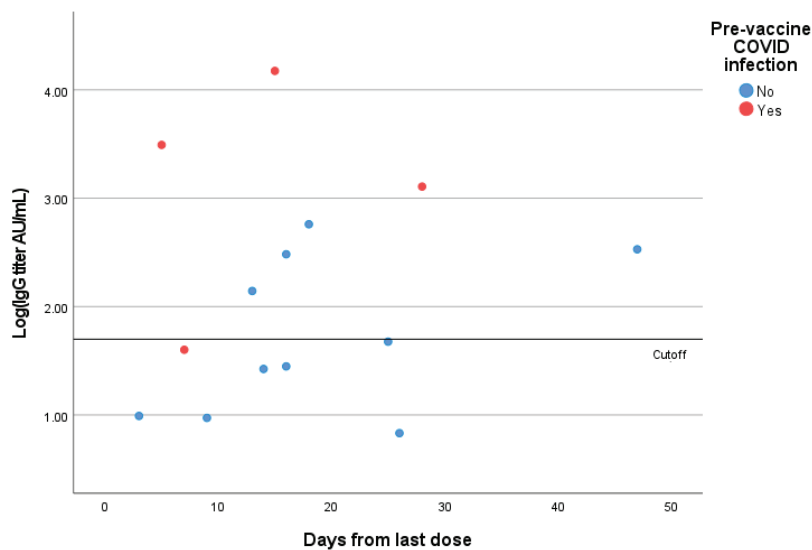


Figure 1. Quantitative IgG titers (logarithmic scale) for patients who received one dose of the vaccine. Each point represents one patient.

Out of 14 patients who have received only one dose of the vaccine, 7 had negative IgG results (seroconversion rate following one dose is 50%). Six out of these 7 patients had no previous COVID-19 infection and only one a prior infection (Figure 1).

Out of 36 patients who have received two doses, six had negative IgG results (17%) and none of them had a prior COVID-19 infection (seroconversion rate following two doses is 83%) (Figure 2). Out of these patients, two were on chemotherapy, one was on chemotherapy and anti-VEGF, two were on Anti-CD 20 therapy and one was taking bispecific T cell engager. One patient with lymphoma on Rituximab had COVID-19 infection 24 days after the second dose of the ChAdOx1 vaccine. This patient had a negative pre-infection IgG test.

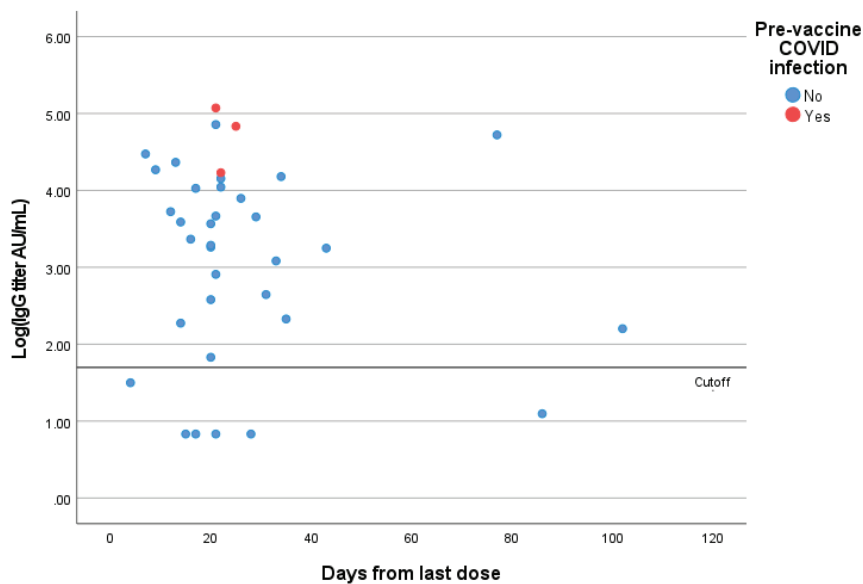


Figure 2. Quantitative IgG titers (logarithmic scale) for patients who received two doses of the vaccine. Each point represents one patient.

Conclusions

To the best of author's knowledge, this is the first report of COVID-19 immune response in cancer patients in a limited resource setting. In our study, seroconversion rates were 50% following the first vaccine dose and 83% after the second dose, slightly lower than data from case series reported from high-income countries [7–10]. A bigger sample size from multiple sources is needed to infer correlations with the type of malignancy, treatment and vaccine type. It is important to note that similar to other studies, patients receiving Anti-CD 20 drugs did not mount an adequate antibody response, likely due to B-cell depleting action. Antibody response is a surrogate marker of vaccine efficacy; however, it does not entirely confer the level of protection provided, part of which is mediated by T-cells [11]. The need to maintain safety precautions post vaccination should be reinforced for all immunosuppressed patients. This is particularly important in low-income countries with lower population vaccination rates compared to high-income countries. Further data from longitudinal studies are required to monitor the response to COVID-19 vaccination in patients with cancer on therapy in different resource settings. Prospectively evaluating the requirement for and efficacy of vaccine booster doses is urgently required.

Funding

This study was funded internally by the Naef K. Basile Cancer Institute. Dr Mukherji is a member of the Cancer and COVID-19 Taskforce and is funded through UK Research and Innovation as part of the Global Challenges Research Fund; Research for Health in Conflict in the Middle East and North Africa (R4HC-MENA) project, grant number ES/P010962/1.

Conflicts of interest

The authors do not report any relevant conflicts of interest.

References

1. *Recommendations of the NCCN COVID-19 Vaccination Advisory Committee (version 3.0)*. [https://www.nccn.org/docs/default-source/covid-19/2021_covid-19_vaccination_guidance_v3-0.pdf?sfvrsn=b483da2b_60] Date accessed: 22/07/21
2. Addeo A, Shah PK, and Bodry N, *et al* (2021) **Immunogenicity of SARS-CoV-2 messenger RNA vaccines in patients with cancer** *Cancer Cell* **39**(8) 1091–1098.e2 <https://doi.org/10.1016/j.ccell.2021.06.009> PMID: [34214473](https://pubmed.ncbi.nlm.nih.gov/34214473/) PMCID: [8218532](https://pubmed.ncbi.nlm.nih.gov/8218532/)
3. Massarweh A, Eliakim-Raz N, and Stemmer A, *et al* (2021) **Evaluation of seropositivity following BNT162b2 messenger RNA vaccination for SARS-CoV-2 in patients undergoing treatment for cancer** *JAMA Oncol* e212155 <https://doi.org/10.1001/jamaoncol.2021.2155>
4. Ehmsen S, Asmussen A, and Jeppesen SS, *et al* (2021) **Antibody and T cell immune responses following mRNA COVID-19 vaccination in patients with cancer** *Cancer Cell* **39**(8) 1034–1036 <https://doi.org/10.1016/j.ccell.2021.07.016> PMID: [34348121](https://pubmed.ncbi.nlm.nih.gov/34348121/) PMCID: [8313483](https://pubmed.ncbi.nlm.nih.gov/8313483/)
5. Yusuf A, Sarfati D, and Booth CM, *et al* (2021) **Cancer and COVID-19 vaccines: a complex global picture** *Lancet Oncol* **22**(6) 749–751 [https://doi.org/10.1016/S1470-2045\(21\)00244-8](https://doi.org/10.1016/S1470-2045(21)00244-8) PMID: [33930324](https://pubmed.ncbi.nlm.nih.gov/33930324/) PMCID: [8078890](https://pubmed.ncbi.nlm.nih.gov/8078890/)
6. Abbott MediaRoom (n.d.). **Abbott receives CE mark for its COVID-19 IgG Quantitative Antibody blood test** [<https://abbott.mediaroom.com/2020-12-15-Abbott-Receives-CE-Mark-for-its-COVID-19-IgG-Quantitative-Antibody-Blood-Test>]
7. Monin L, Laing AG, and Muñoz-Ruiz M, *et al* (2021) **Safety and immunogenicity of one versus two doses of the COVID-19 vaccine BNT162b2 for patients with cancer: interim analysis of a prospective observational study** *Lancet Oncol* **22**(6) 765–778 [https://doi.org/10.1016/S1470-2045\(21\)00213-8](https://doi.org/10.1016/S1470-2045(21)00213-8) PMID: [33930323](https://pubmed.ncbi.nlm.nih.gov/33930323/) PMCID: [8078907](https://pubmed.ncbi.nlm.nih.gov/8078907/)
8. Palich R, Veyri M, and Solas C, *et al* (2021) **High seroconversion rate but low antibody titers after two injections of BNT162b2 (Pfizer-BioNTech) vaccine in patients treated by chemotherapy for solid cancers** *Ann Oncol* <https://doi.org/10.1016/j.annonc.2021.06.018>
9. Goshen-Lago T, Waldhorn I, and Holland R, *et al* (2021) **Serologic status and toxic effects of the SARS-CoV-2 BNT162b2 vaccine in patients undergoing treatment for cancer** *JAMA Oncol* e212675 <https://doi.org/10.1001/jamaoncol.2021.2675> PMID: [34236381](https://pubmed.ncbi.nlm.nih.gov/34236381/) PMCID: [8267843](https://pubmed.ncbi.nlm.nih.gov/8267843/)
10. Thakkar A, Gonzalez-Lugo JD, and Goradia N, *et al* (2021) **Seroconversion rates following COVID-19 vaccination among patients with cancer** *Cancer Cell* **39**(8) 1081–1090.e2 <https://doi.org/10.1016/j.ccell.2021.06.002> PMID: [34133951](https://pubmed.ncbi.nlm.nih.gov/34133951/) PMCID: [8179248](https://pubmed.ncbi.nlm.nih.gov/8179248/)
11. Poland GA, Ovsyannikova IG, and Kennedy RB (2020) **SARS-CoV-2 immunity: review and applications to phase 3 vaccine candidates** *Lancet* **396**(10262) 1595–1606 [https://doi.org/10.1016/S0140-6736\(20\)32137-1](https://doi.org/10.1016/S0140-6736(20)32137-1) PMID: [33065034](https://pubmed.ncbi.nlm.nih.gov/33065034/) PMCID: [7553736](https://pubmed.ncbi.nlm.nih.gov/7553736/)